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Are opportunistic captures of neonate ungulates biasing relative estimates of litter size?

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Abstract

The capture of neonate ungulates allows for the collection of valuable ecological data, including estimates of litter size. However, varied methods used to capture neonate ungulates can result in sampling biases. Our objective was to determine if opportunistic captures of neonate ungulates (i.e., locating neonates by visually scanning for adult females displaying postpartum behaviors) bias relative estimates of litter size and investigate potential causes if a bias does exist. We analyzed data from 161 litters of mule deer (*Odocoileus hemionus*) sampled using three different capture methods during 2019–2021 in Utah, USA. Estimates of litter size derived from opportunistic captures were smaller than estimates derived from movement-based captures or captures completed with the aid of vaginal implant transmitters (VITs). Age at capture was inversely related to estimates of litter size and likely influenced the detection bias associated with opportunistic captures. Neonates captured opportunistically were not older than neonates captured using movement-based methods, but were older than neonates captured with the aid of VITs. Distance between neonates from the same litter did not influence estimates of litter size. Researchers should be aware of the biases associated with different capture methods and use caution when interpreting data among multiple capture methods. Estimates of litter size derived from opportunistic captures should not be compared to estimates of litter size derived from alternative capture methods without accounting for the detection bias we observed.

Keywords: Capture methods, Litter size, Mule deer, Neonates, *Odocoileus hemionus*, Sampling bias, Ungulates, Utah, Vaginal implant transmitters

Background

Multiple methods have been utilized to locate neonate ungulates and improve the likelihood of capture. Neonates can be captured opportunistically, without prior knowledge of a specific parturition event, by searching potential parturition sites or observing postpartum behaviors by adult females [1–6]. Opportunistic captures can also be performed using aerial/terrestrial vehicles,

audio recordings, spotlights, thermal sensors, or trained animals [7–12]. Alternatively, neonate ungulates can be captured with prior knowledge of a specific parturition event. Parturient ungulates often display characteristic movement patterns, so targeted captures can occur by monitoring the movement of adult females with tracking collars [13–15]. Targeted captures can also occur with the aid of vaginal implant transmitters (VITs), a radio transmitter expelled at parturition that signals the occurrence of a parturition event [16–18]. Capturing neonate ungulates allows for the collection of valuable ecological data, including early-life survival, cause-specific mortality,

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morphometry, movement, parturition date/location, and sex ratio.

Notably, the variety of methods utilized to capture neonate ungulates has led to concerns of sampling biases. For example, age at capture tends to be higher for neonates captured using opportunistic methods than for neonates captured with the aid of VITs [19, 20]. Opportunistic captures are also more likely than VIT-aided captures to miss early-life mortalities, biasing estimates of survival and cause-specific mortality [18, 21-23]. Moreover, opportunistic captures may not be randomly distributed, resulting in biased distributions of parturition events favoring researcher access [16, 24, 25]. An additional, untested bias when capturing neonate ungulates may be skewed estimates of litter size. This knowledge gap is problematic considering litter size is frequently estimated during early-life capture events [26–31]. While estimates of litter size derived from early-life capture events are likely prone to detection failure, relative estimates can still be utilized to make comparisons among years, sites, and age classes.

Litter size has major implications for individual fitness and population dynamics in species with variable litter sizes. For example, litter size within a species tends to be positively related to the total body mass or relative body fat of female ungulates [32-35]. Following parturition, female ungulates with larger litters incur greater lactation costs and have reduced fat reserves compared to conspecifics with smaller litters [36–38]. The implications of litter size also extend to neonate ungulates. Neonates in larger litters tend to have lower rates of survival than conspecifics in smaller litters [27-29]. Neonate mass within a species is also inversely related to litter size [33, 39, 40]. At the population level, rates of recruitment may increase with larger litter sizes [27]. Despite the potential ecological implications of litter size, the potential biases associated with method of capture when estimating litter size of ungulates are unknown.

Our objective was to determine if opportunistic captures of neonate ungulates bias relative estimates of litter size and investigate potential causes if a bias does exist. We used captures of neonate mule deer (*Odocoileus hemionus*) and three capture methods (opportunistic, movement-based, and VIT-aided) as an initial case study. Because opportunistic methods tend to capture older individuals than alternative methods and bias estimates of survival, we hypothesized that opportunistic captures would also bias relative estimates of litter size [18–23]. We predicted that estimates of litter size derived from opportunistic captures would be smaller than estimates of litter size derived from movement-based or VIT-aided captures. We also hypothesized that age at capture and distance between neonates from the same litter would

be linked to estimates of litter size and capture method. Likelihood of capture tends to decrease as neonates age, and neonates from the same litter often bed separately once moved from the parturition site [41–44]. Thus, we predicted that new hoof growth (a proxy for age) would be inversely related to litter size and would be greater for litters associated with opportunistic captures than for litters associated with movement-based or VIT-aided captures. Further, we predicted that distance between neonates from the same litter would be inversely related to new hoof growth and would be greater for litters associated with opportunistic captures than for litters associated with movement-based or VIT-aided captures.

Methods

Study area

We performed this study in the Book Cliffs (39.5°, -109.3°) and the Cache (41.7°, -111.5°) management units of Utah, USA (Fig. 1). Elevations in the Book Cliffs spanned from approximately 1675 m to 2590 m, average annual precipitation was 22.9 cm, and average annual temperature was 10.3 °C (averages from 1981 to 2010) [45]. Terrain in the Book Cliffs consisted of cliff faces,

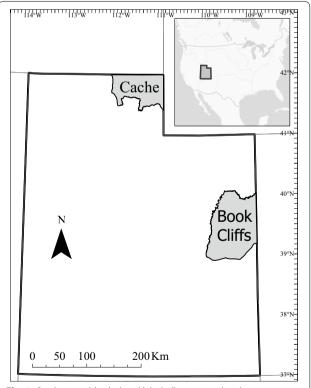


Fig. 1 Study areas (shaded and labeled) associated with captures of neonate mule deer during 2019–2021 in Utah, USA. A total of 128 neonates were captured in the Book Cliffs management unit and 110 neonates were captured in the Cache management unit

ridges, valleys, and flatlands over an area of proximately 9300 km². The vegetation community in the Book Cliffs varied from a sagebrush steppe (Artemisia spp.) to a pine-juniper woodland (Pinus monophyla-Juniperus oteosperma). Elevations in the Cache spanned from approximately 1300 m to 3040 m, average annual precipitation was 43.4 cm, and average annual temperature was 5.1 °C (averages from 1981 to 2010) [45]. Terrain in the Cache consisted of ridges and valleys over an area of approximately 4000 km². The vegetation community in the Cache varied from a sagebrush steppe to a pineaspen woodland (Pseudotsuga menziesii-Populus tremuloides). Predators of mule deer in the Book Cliffs and Cache included black bears (Ursus americanus), bobcats (Lynx rufus), coyotes (Canis latrans), and mountain lions (Puma concolor).

Adult captures

An independent capture company (Helicopter Wildlife Services, Austin, TX, USA or Quicksilver Air Inc., Peyton, CO, USA) captured adult (i.e., ≥ 2 years old) female mule deer via helicopter net-gunning during February-March of 2019-2021 [46, 47]. Following capture, the capture company hobbled and blindfolded animals to minimize stress and transported animals to a nearby processing site. At the processing site, we fitted animals with a tracking collar (G5-2DH, 595g, Advanced Telemetry Systems, Isanti, MN, USA) which included global positioning system (GPS) technology that recorded coordinates every 2 h. Collars were also equipped with a very high frequency (VHF) transmitter. In addition, we checked the pregnancy status of animals via transabdominal ultrasonography [16, 48]. If an animal was pregnant, we inserted a VIT (M3930U, 23g, Advanced Telemetry Systems) using a vaginoscope [16, 24]. All VITs included light and temperature sensors to detect expulsion, and a VHF transmitter. An ultra high frequency (UHF) link between each collar and VIT allowed for near-instant notification of VIT expulsion, sent via email [49].

Neonate captures

We captured neonate mule deer during May–July of 2019–2021 using 3 capture methods: opportunistic, movement-based, and VIT-aided. To perform opportunistic captures, we visually scanned potential mule deer habitat for adult females [50, 51]. If females displayed typical postpartum behaviors (e.g., increased vigilance, reluctance to flee, social isolation, or vocalizations) or physical characteristics (e.g., enlarged udder or sunken flanks), we continued monitoring the female for evidence of neonates [50–52]. Upon observation of a neonate, we proceeded directly to the neonate's location. If we did not

observe a neonate, we systematically searched the area surrounding the female's location.

To perform movement-based captures, we monitored movement patterns of adult females with tracking collars, but without a VIT [43, 53]. If females displayed movement patterns indicative of parturition (e.g., a sudden, lengthy movement followed by a sustained reduction and localization of movement), we systematically searched the area surrounding the female's most recent coordinates [43, 54, 55]. The sample of collared females without a VIT included animals that previously expelled a VIT (and were incorporated into the study during a previous year) and animals that were not pregnant at the time of capture (but became pregnant during a subsequent year).

To perform VIT-aided captures, we systematically searched the area surrounding expelled VITs [16, 52, 56]. Generally, we waited more than 3 h after VIT expulsion before initiating a search to allow for female—neonate bonding to occur [56–58]. Wait times varied based on accessibility of parturition sites and the number of parturition events that occurred on the same day. If neonates were not located near the parturition site, we searched the female's most recent coordinates.

Data collection

Following capture of a neonate, we blindfolded the animal and completed processing while wearing nitrile gloves. We recorded GPS coordinates of the capture location and fitted each neonate with an expandable tracking collar (M4230BU, 125g, Advanced Telemetry Systems). We also recorded new hoof growth—the distance from the hairline to the distal edge of the growth ring on a front hoof [59-61]. Processing time was typically < 10 min. If a lone neonate was found, we began searching the surrounding area for additional neonates from the same litter. All similar-sized neonates located in the same search area during the same search period were assumed to be from the same litter. We confirmed the validity of this assumption using a UHF link between neonate collars and adult collars worn by mothers associated with movement-based or VIT-aided captures. We received notification via email if collared mothers and their offspring were in close proximity, which indicated that mother-offspring relationships were correctly identified. Although the mother-offspring relationships of neonates without a collared mother could not be confirmed post-capture, we assumed that our mother-offspring identification was consistent among capture methods. We estimated litter size based on the number of neonates located during our search, even if neonates fled before capture could occur. If it became too dark to locate neonates, we postponed searches until the next day.

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Data analysis

To determine if opportunistic captures biased estimates of litter size, and why any bias may have occurred, we analyzed capture data and estimates of litter size using 2 logistic regression models and 3 linear regression models. We did not combine models into a multivariate regression framework because explanatory variables showed evidence of correlation [62, 63]. When creating models that examined the relationship of age at capture, we used new hoof growth as a proxy for age at capture. New hoof growth is linearly related to the age of neonate mule deer and has been used to estimate age when exact birth dates are not known [21, 30, 33, 64]. However, established equations to estimate age can produce ages that vary by more than 11 days [57, 60]. Similar equations established for white-tailed deer (Odocoileus virginianus) can produce estimated ages that are accurate (i.e., within 1-3 days of the true age) < 50% of the time [61]. Thus, we did not convert new hoof growth measurements to an estimated age at capture to avoid cases of pseudo-accuracy.

We used a mixed-effects logistic regression to examine the relationship between capture method and litter size (coded as litter size of 1=0, litter size of 2=1). We also used a mixed-effects logistic regression to examine the relationship between mean new hoof growth per litter (henceforth shortened to "new hoof growth") and litter size. We used a mixed-effects linear regression to examine the relationship between capture method and new hoof growth. We also used a mixed-effects linear regression to examine the relationship between new hoof growth and distance between neonates from the same litter. Finally, we used a mixed-effects linear regression to examine the relationship between capture method and distance between neonates from the same litter. To limit confounding variables that might influence hoof growth measurements and/or distances between neonates from the same litter, we censored neonates that were found underdeveloped/stillborn (n=12), predated prior to our arrival (n=2), or captured on a different date than neonates from the same litter (n=2). In addition, we censored neonates that fled before data could be collected (n=3). In all models, we included management unit and year as random effects and confirmed that relevant assumptions were met [62, 63]. We analyzed data using the lme4 package in Program R version 4.0.2 and used an α -value of 0.05 in all interpretations [65, 66].

Results

We analyzed data from 161 litters (238 neonates) of mule deer captured over a period of 3 years. Of the litters we analyzed, 90 litters (128 neonates) were associated with the Book Cliffs management unit and 71 litters (110 neonates) were associated with the Cache management unit. We located 64 litters using opportunistic methods, 14 litters using movement-based methods, and 83 litters using VIT-aided methods. Based on our estimates of litter size, 84 litters (52.2%) comprised 1 neonate and 77 litters (47.8%) comprised 2 neonates. We did not find evidence of any litters comprising more than 2 neonates. Mean (\pm SE) litter size derived from captures on the Book Cliffs (1.4 \pm 0.1) and the Cache (1.5 \pm 0.1) were not different (P=0.11). There was no evidence that any mother–offspring relationships were incorrectly identified.

Estimates of litter size derived from opportunistic captures were smaller than estimates of litter size derived from movement-based (β =1.3, OR=3.7, 95% CI=1.1–12.7, SE=0.6, z_{11} =2.1, P=0.03) or VIT-aided captures (β =1.5, OR=4.6, 95% CI=2.2–9.6, SE=0.4, z_{80} =4.2, P<0.001). Estimates of mean (\pm SE) litter size derived from opportunistic captures, movement-based captures, and VIT-aided captures were 1.3 \pm 0.1 neonates, 1.6 \pm 0.1 neonates, and 1.6 \pm 0.1 neonates, respectively (Fig. 2). There was an inverse relationship between new hoof growth and estimated litter size (β = -0.3, SE=0.1, z_{152} = -2.0, P=0.04; Fig. 3). New hoof growth associated with opportunistic captures was not different than new hoof growth associated with movement-based captures (β = -0.3, SE=0.4, t_{10} = -0.7, P=0.46, R^2m =0.1,

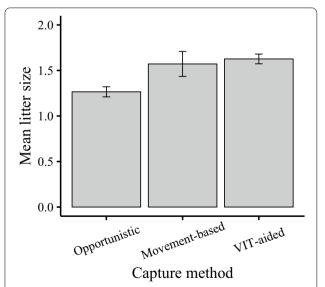


Fig. 2 Estimates of mean litter size derived from captures of neonate mule deer during 2019–2021 in Utah, USA. Capture methods included visually scanning for females displaying postpartum behaviors (opportunistic), monitoring movement patterns of females with tracking collars (movement-based), and searching the area surrounding expelled vaginal implant transmitters (VITs; VIT-aided). Error bars represent standard error

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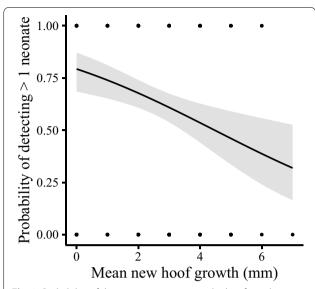


Fig. 3 Probability of detecting > 1 neonate mule deer from the same litter in relation to mean new hoof growth (a proxy for neonate age) once a litter had been detected. The shaded area represents a 95% confidence interval. All litters (*n* = 153) were detected during 2019–2021 in Utah, USA

 $R^2c = 0.3$), but was greater than new hoof growth associated with VIT-aided captures ($\beta = -1.1$, SE = 0.3, $t_{73} = -4.0$, $P \le 0.001$, $R^2 m = 0.1$, $R^2 c = 0.3$). New hoof growth means (\pm SE) associated with opportunistic captures, movement-based captures, and VIT-aided captures were 3.3 ± 0.2 mm, 2.9 ± 0.3 mm, and 1.8 ± 0.2 mm, respectively (Fig. 4). There was no relationship between new hoof growth and distance between neonates from the same litter ($\beta = 2.2$, SE = 1.3, $t_{64} = 1.7$, P = 0.10). Distances between neonates from the same litter associated with opportunistic captures were shorter than distances associated with movement-based captures ($\beta = 1.4$, SE=0.6, t_5 =2.4, P=0.02, R^2m =0.1, R^2c =0.2), but were not different than distances associated with VIT-aided captures ($\beta = 0.9$, SE = 0.5, $t_{43} = 1.9$, P = 0.06, $R^2 m = 0.1$, $R^2c = 0.2$). Mean (\pm SE) distance between neonates from the same litter associated with opportunistic captures, movement-based captures, and VIT-aided captures was 1.7 ± 1.1 m, 21.9 ± 12.2 m, and 5.7 ± 1.2 m, respectively (Fig. 5).

Discussion

Our results indicate that estimates of litter size derived from opportunistic captures were smaller than estimates of litter size derived from movement-based or VIT-aided captures. Interpretations of estimates of litter size derived from opportunistic captures should not be made without accounting for this bias. While all capture methods likely had some level of detection failure when

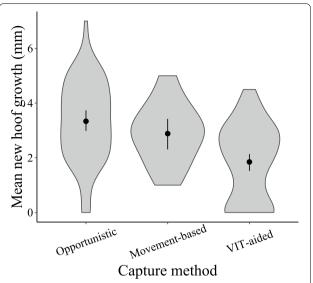


Fig. 4 Mean new hoof growth per litter (a proxy for neonate age) associated with captures of neonate mule deer during 2019–2021 in Utah, USA. Capture methods included visually scanning for females displaying postpartum behaviors (opportunistic), monitoring movement patterns of females with tracking collars (movement-based), and searching the area surrounding expelled vaginal implant transmitters (VITs; VIT-aided). Bars represent 95% confidence intervals

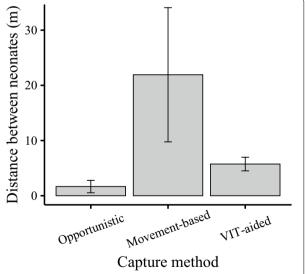


Fig. 5 Mean distances between neonates from the same litter associated with captures of neonate mule deer during 2019–2021 in Utah, USA. Capture methods included visually scanning for females displaying postpartum behaviors (opportunistic), monitoring movement patterns of females with tracking collars (movement-based), and searching the area surrounding expelled vaginal implant transmitters (VITs; VIT-aided). Error bars represent standard error

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estimating litter size, opportunistic captures appear to increase the likelihood of detection failure. The only in utero evaluation of mule deer litter size that has occurred in Utah (as far as we are aware) reported a mean litter size of 1.8 for adults ≥ 2 years old [67]. Our estimates of mean litter size derived from movement-based and VITaided captures were closer to the previous estimate than our estimate of mean litter size derived from opportunistic captures (x = 1.6, 1.6, and 1.3, respectively). Sampling biases associated with opportunistic captures are well recorded, especially when compared to VIT-aided captures [18-23]. Potential sampling biases using movement-based captures, however, are poorly understood. While our study provides one example of movementbased methods minimizing bias when capturing neonate ungulates, our sample size for litters associated with movement-based captures was relatively small (n = 14). We present interpretations of our results associated with movement-based captures not as concrete conclusions, but as potential patterns to inspire future research [68]. Movement-based captures may be utilized more often as fine-scale movement data increases in availability and statistical analyses to detect parturition improve [43, 53].

Our investigation into why estimates of litter size were smaller for opportunistic captures revealed that age at capture was an influential factor. As new hoof growth (which served as a proxy for age at capture) increased, the probability of detecting >1 neonate from the same litter decreased. This inverse relationship between age at capture and probability of detection was likely due to a combination of increased neonate mobility and the occurrence of mortality events. The likelihood of capturing neonate mule deer decreases when searches are initiated ≥ 4 days post-parturition, and mean age at capture using opportunistic methods generally ranges between 3 and 7 days [21, 43, 51, 69]. In addition, the age difference between neonates captured using VITs and neonates captured opportunistically is large to enough to result in differential estimates of survival [18, 21-23]. Researchers should keep in mind species-specific precocity and risk of early-life mortality when attempting to estimate litter sizes from other ungulates. Interestingly, new hoof growths associated with opportunistic and movementbased captures did not differ despite opportunistic and movement-based captures producing different estimates of litter size. This result suggests age at capture is not the only factor that influences estimates of litter size, and an additional difference in capture methodology may have even greater influence. An increased awareness of the recent locations of female ungulates may allow for movement-based captures to more effectively locate neonates from the same litter, but more work is required to confirm this suspicion.

We did not find strong evidence supporting the idea that differences in distance between neonates from the same litter resulted in opportunistic captures biasing estimates of litter size. Age at capture was not linked to distance between neonates from the same litter. Distances between neonates from the same litter associated with opportunistic captures were shorter than distances associated with movement-based captures, but were not different than distances associated with VIT-aided captures. However, the increased detection bias associated with opportunistic captures may have influenced our results. Female ungulates often separate neonates soon after parturition [41, 42, 50, 70]. Distance of separation varies, but can be more than 400 m for mule deer [70]. Thus, when performing opportunistic captures we may have failed to detect multiple neonates from the same litter unless neonates were close together. While we attempted to exercise equal effort in all searches, sampling protocols that dictated a standardized search distance may have helped to minimize sampling bias [51, 52]. A relatively standardized search distance for movement-based captures (the area surrounding the female's most recent coordinates) may have resulted in movement-based captures having greater distances between neonates from the same litter.

Conclusions

Capturing neonate mule deer using opportunistic methods results in smaller (and likely less accurate) estimates of litter size compared to movement-based and VIT-aided captures. An increased age at capture appears to be one of the primary reasons why this detection bias exists. As litter size continues to be utilized as an indicator of individual fitness and population trends, researchers should be aware of the biases of using different methods to capture neonate ungulates. We suggest using movement-based or VIT-aided captures to ensure minimal bias when estimating litter size during early-life capture events.

Abbreviations

VIT: Vaginal implant transmitter; GPS: Global positioning system; VHF: Very high frequency; UHF: Ultra high frequency.

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Author contributions

BM, RL, and KH conceived the study design and acquired funding. All authors contributed to data collection. MT, BM, RL, and TH analyzed and interpreted data. MT drafted the manuscript with substantial revisions from all authors. All agreed to be accountable for the accuracy and integrity of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

All methods were approved by the Institutional Animal Care and Use Committee at Brigham Young University (protocol 19-0202) and followed guidelines from the American Society of Mammalogists [71].

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Ballard WB, Whitlaw HA, Young SJ, Jenkins RA, Forbes GJ. Predation and survival of white tailed deer fawns in northcentral New Brunswick. J Wild Manag. 1999;63:574–9.
- Pettorelli N, Gaillard J-M, Yoccoz NG, Duncan P, Maillard D, Delorme D, et al. The response of fawn survival to changes in habitat quality varies according to cohort quality and spatial scale. J Anim Ecol. 2005;74:972–81.
- 3. Hiller TL, Campa H, Winterstein SR, Rudolph BA. Survival and space use of fawn white-tailed deer in southern Michigan. Am Midl Nat. 2008;159:403–12.
- Grovenburg TW, Jacques CN, Klaver RW, Jenks JA. Bed site selection by neonate white-tailed deer in grassland habitats on the northern Great Plains. J Wild Manag. 2010;2010(74):1250–6.
- Hurley MA, Unsworth JW, Zager P, Hebblewhite M, Garton EO, Montgomery DM, et al. Demographic response of mule deer to experimental reduction of coyotes and mountain lions in southeastern Idaho. Wildl Monogr. 2011;178:1–33.
- Pitman JW, Cain JW III, Liley SG, Gould WR, Quintana NT, Ballard WB. Post-parturition habitat selection by elk calves and adult female elk in New Mexico. J Wild Manag. 2014;78:1216–27.
- Singer FJ, Harting A, Symonds KK, Coughenour MB. Density dependence, compensation, and environmental effects on elk young mortality in Yellowstone National Park. J Wild Manag. 1997;61:12–25.
- Vreeland JK, Diefenbach DR, Wallingford BD. Survival rates, mortality causes, and habitats of Pennsylvania white-tailed deer fawns. Wildl Soc Bull. 2004;32:542–53.
- Ditchkoff SS, Raglin JB, Smith JM, Collier BA. From the field: capture of white-tailed deer fawns using thermal imaging technology. Wildl Soc Bull. 2005;33:1164–8.
- Smith WP, Coblentz BE. Cattle or sheep reduce fawning habitat available to Columbian white-tailed deer in western Oregon. Northwest Sci. 2010;84:315–26.
- Hasapes SK, Comer CE. White-tailed deer fawn survival, home range, and habitat composition in northwest Louisiana. Wildl Soc Bull. 2017;41:499–507.

- Obermoller TR, Norton AS, Michel ES, Haroldson BS. Use of drones with thermal infrared to locate white-tailed deer neonates for capture. Wildl Soc Bull. 2021;45:682–9.
- 13. Kunkel KE, Mech LD. Wolf and bear predation on white-tailed deer and fawns in north-eastern Minnesota. Can J Zool. 1994;72:1557–65.
- DelGiudice GD, Severud WJ, Obermoller TR, Wright RG, Enright TA, St-Louis V. Monitoring movement behavior enhances recognition and understanding of capture-induced abandonment of moose neonates. J Mammal. 2015;96:1005–16.
- Severud WJ, Obermoller TR, DelGiudice GD, Fieberg JR. Survival and cause-specific mortality of moose calves in northeastern Minnesota. J Wild Manag. 2019;83:1131–42.
- Bishop CJ, Freddy DJ, White GC, Watkins BE, Stephenson TR, Wolfe LL. Using vaginal implant transmitters to aid in capture of mule deer neonates. J Wild Manag. 2007;71:945–54.
- 17. Rearden SN, Anthony RG, Johnson BK. Birth-site selection and predation risk of Rocky Mountain elk. J Mammal. 2011;92:1118–26.
- 18. Dion JR, Haus JM, Rogerson JE, Bowman JL. White-tailed deer neonate survival in the absence of predators. Ecosphere. 2020;11: e03122.
- Nelson MA, Cherry MJ, Howze MB, Warren RJ, Conner LM. Coyote and bobcat predation on white-tailed deer fawns in a longleaf pine ecosystem in southwestern Georgia. J Southeast Assoc Fish Wildl Agencies. 2015;2:208–13.
- Kautz TM, Belant JL, Beyer DE Jr, Strickland BK, Petroelje TR, Sollmann R. Predator densities and white-tailed deer fawn survival. J Wild Manag. 2019;83:1261–70
- Gilbert SL, Lindberg MS, Hundertmark KJ, Person DK. Dead before detection: addressing the effects of left truncation on survival estimation and ecological inference for neonates. Methods Ecol Evol. 2014;5:992–1001.
- Chitwood MC, Lashley MA, DePerno CS, Moorman CE. Considerations on neonatal ungulate capture method: potential for bias in survival estimation and cause specific mortality. Wildl Biol. 2017. https://doi.org/10.2981/ wlb.00250
- Brackel KL, Michel ES, Gullikson BS, Jenks JA, Jensen WF. Capture method affects survival estimates and subsequent interpretation of ecological covariates for a long-lived cervid. Ecol Evol. 2021;11:6444–55.
- Bishop CJ, Anderson CR, Walsh DP, Bergman EJ, Kuechle P, Roth J. Effectiveness of a redesigned vaginal implant transmitter in mule deer. J Wild Manag. 2011;75:1797–806.
- Kilgo JC, Ray HS, Vukovich M, Goode MJ, Ruth C. Predation by coyotes on white-tailed deer neonates in South Carolina. J Wild Manag. 2012;76:1420–30.
- Lomas LA, Bender LC. Survival and cause-specific mortality of neonatal mule deer fawns, north-central New Mexico. J Wild Manag. 2007;71:884–94.
- 27. Johnstone-Yellin TL, Shipley LA, Myers WL, Robinson HS. To twin or not to twin? Trade-offs in litter size and fawn survival in mule deer. J Mammal. 2009;90:453–60.
- Keech MA, Lindberg MS, Boertje RD, Valkenburg P, Taras BD, Boudreau TA, et al. Effects of predator treatments, individual traits, and environment on moose survival in Alaska. J Wild Manag. 2011;75:1361–80.
- Chitwood MC, Lashley MA, Kilgo JC, Pollock KH, Moorman CE, DePerno CS. Do biological and bedsite characteristics influence survival of neonatal white-tailed deer? PLoS ONE. 2015;10: e0119070.
- 30. Shallow JRT, Hurley MA, Monteith KL, Bowyer RT. Cascading effects of habitat on maternal condition and life-history characteristics of neonatal mule deer. J Mammal. 2015;96:194–205.
- 31. Shuman RM, Cherry MJ, Simoneaux TN, Dutoit EA, Kilgo JC, Chamberlain MJ, et al. Survival of white-tailed deer neonates in Louisiana. J Wild Manag. 2017;81:834–45.
- Keech MA, Bowyer RT, Ver Hoef JM, Boertje RD, Dale BW, Stephenson TR. Life-history consequences of maternal condition in Alaskan moose. J Wild Manag. 2000;64:450–62.
- 33. Monteith KL, Bleich VC, Stephenson TR, Pierce BM, Conner MM, Kie JG, et al. Life-history characteristics of mule deer: effects of nutrition in a variable environment. Wildl Monogr. 2014;186:1–62.
- Frauendorf M, Gethöffer F, Siebert U, Keuling O. The influence of environmental and physiological factors on the litter size of wild boar (Sus scrofa) in an agriculture dominated area in Germany. Sci Total Environ. 2016;541:877–82.

- 35. Flajšman K, Borowik T, Pokorny B, Jędrzejewska B. Effects of population density and female body mass on litter size in European roe deer at a continental scale. Mam Res. 2018;2018(63):91–8.
- 36. Carl GR, Robbins CT. The energetic cost of predator avoidance in ungulates: hiding versus following. Can J Zool. 1988;66:239–46.
- Monteith KL, Stephenson TR, Bleich VC, Conner MM, Pierce BM, Bowyer RT. Risk-sensitive allocation in seasonal dynamics of fat and protein reserves in a long-lived mammal. J Anim Ecol. 2013;82:377–88.
- Simard MA, Huot J, de Bellefeuille S, Côté SD. Linking conception and weaning success with environmental variation and female body condition in a northern ungulate. J Mammal. 2014;95:311–27.
- Sæther BE, Heim M. Écological correlates of individual variation in age at maturity in female moose (*Alces alces*): the effects of environmental variability. J Anim Ecol. 1993;62:482–9.
- Michel ES, Demarais S, Strickland BK, Belant JL. Contrasting the effects of maternal and behavioral characteristics on fawn birth mass in whitetailed deer. PLoS ONE. 2015;10: e0136034.
- 41. Barrett MW. Movements, habitat use, and predation on pronghorn fawns in Alberta. J Wild Manag. 1984;48:542–50.
- 42. Blank DA, Ruckstuhl K, Yang W. Antipredator strategy of female goitered gazelles (*Gazella subgutturosa* Guld., 1780) with hiding fawn. Behav Process. 2015;119:44–9.
- Peterson ME, Anderson CR Jr, Alldredge WM, Doherty PF Jr. Using maternal mule deer movements to estimate timing of parturition and assist fawn captures. Wildl Soc Bull. 2018;42:616–21.
- Turnley MT, Larsen RT, Hersey KR, Sallee DW, Hinton MS, McMillan BR.
 Optimizing methods for capturing neonate elk when using vaginal implant transmitters. Wild Soc Bull. 2022;46: e1326.
- Western Regional Climate Center. Cooperative climatological data summaries. https://wrcc.dri.edu/Climate/west_coop_summaries.php. Accessed 14 Dec 2021.
- 46. Barrett MW, Nolan JW, Roy LD. Evaluation of a hand-held net-gun to capture large mammals. Wild Soc Bull. 1982;10:108–14.
- Krausman PR, Hervert JJ, Ordway LL. Capturing deer and mountain sheep with a net-gun. Wild Soc Bull. 1985;13:71–3.
- Stephenson TR, Testa JW, Adams GP, Sasser RG, Schwartz CC, Hundertmark KJ. Diagnosis of pregnancy and twinning in moose by ultrasonography and serum assay. Alces. 1995;31:167–72.
- Dion JR, Haus JM, Rogerson JE, Bowman JL. An initial performance review of vaginal implant transmitters paired with GPS collars. Anim Biotelem. 2019;7:1–6.
- White M, Knowlton FF, Glazener WC. Effects of dam-newborn fawn behavior on capture and mortality. J Wild Manag. 1972;36:897–906.
- 51. Pojar TM, Bowden DC. Neonatal mule deer fawn survival in west-central Colorado. J Wild Manag. 2004;68:550–60.
- 52. Carstensen M, DelGiudice GD, Sampson BA. Using doe behavior and vaginal-implant transmitters to capture neonate white-tailed deer in northcentral Minnesota. Wild Soc Bull. 2003;31:634–41.
- Nicholson KL, Warren MJ, Rostan C, Mansson J, Paragi TF, Sand H. Using fine-scale movement patterns to infer ungulate parturition. Ecol Indic. 2019;101:22–30.
- 54. Vore JM, Schmidt EM. Movements of female elk during calving season in northwest Montana. Wild Soc Bull. 2001;29:720–5.
- Long RA, Kie JG, Bowyer RT, Hurley MA. Resource selection and movements by female mule deer *Odocoileus hemionus*: effects of reproductive stage. Wildl Biol. 2009;15:288–98.
- Quintana NT, Ballard WB, Wallace MC, Krausman PR, deVos J Jr, Alcumbrac O, et al. Survival of mule deer fawns in central Arizona. Southwest Nat. 2016;61:93–100.
- Haskell SP, Ballard WB, Butler DA, Tatman NM, Wallace MC, Kochanny CO, et al. Observations on capturing and aging deer fawns. J Mammal. 2007:88:1482–7
- Tatman NM, Ballard WB, Wallace MC, Haskell SP, Krausman PR, deVos J Jr, et al. Evaluation of use of vaginal-implant transmitters in mule deer (Odocoileus hemionus). Southwest Nat. 2011;56:247–51.
- 59. Haugen AO, Speake DW. Determining the age of young fawn white-tailed deer. J Wild Manag. 1958;22:319–21.
- Robinette WL, Baer CH, Pillmore RE, Knittle CE. Effects of nutritional change on captive mule deer. J Wild Manag. 1973;37:312–26.

- Sams MG, Lochmiller RL, Hellgren EC, Warde WD, Varner LW. Morphometric predictors of neonatal age for white-tailed deer. Wild Soc Bull. 1996:24:53–7
- Stoltzfus JC. Logistic regression: a brief primer. Acad Emerg Med. 2011;18:1099–104.
- Casson RJ, Farmer LDM. Understanding and checking the assumptions of linear regression: a primer for medical researchers. Clin Exp Ophthalmol. 2014;42:590–6.
- 64. Whittaker DG, Lindzey FG. Effect of coyote predation on early fawn survival in sympatric deer species. Wild Soc Bull. 1999;27:256–62.
- 65. Bates D, Mächler M, Bolker B, Walker S. Fitting linear mixed-effects models using Ime4. J Stat Softw. 2015;67:1.
- R Core Team. R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2020. http://www.r-project.org.
- 67. Robinette WL, Gashwiler JS, Jones DA, Crane HS. Fertility of mule deer in Utah. J Wild Manag. 1955;19:115–36.
- Bissonette JA. Small sample size problems in wildlife ecology: contingent analytical approach. Wildl Biol. 1999;5:65–71.
- 69. Steigers WD, Flinders JT. Mortality and movements of mule deer fawns in Washington. J Wild Manag. 1980;1980(44):381–8.
- Riley SJ, Dood AR. Summer movements, home range, habitat use, and behavior of mule deer fawns. J Wild Manag. 1984;48:1302–10.
- Sikes RS, Animal Care and Use Committee of the American Society of Mammalogists. 2016 guidelines of the American Society of Mammalogists for the use of wild mammals in research and education. J Mammal. 2016;97:663–88.

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